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EXAMINER

POHNERT, STEVEN C

ART UNIT	PAPER NUMBER
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1634

NOTIFICATION DATE	DELIVERY MODE
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02/03/2009

ELECTRONIC

Please find below and/or attached an Office communication concerning this application or proceeding.

The time period for reply, if any, is set in the attached communication.

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Office Action Summary	Application No. 10/535,442	Applicant(s) ROTH ET AL.	
	Examiner Steven C. Pohnert	Art Unit 1634	

-- The MAILING DATE of this communication appears on the cover sheet with the correspondence address --

Period for Reply

A SHORTENED STATUTORY PERIOD FOR REPLY IS SET TO EXPIRE 3 MONTH(S) OR THIRTY (30) DAYS, WHICHEVER IS LONGER, FROM THE MAILING DATE OF THIS COMMUNICATION.

- Extensions of time may be available under the provisions of 37 CFR 1.136(a). In no event, however, may a reply be timely filed after SIX (6) MONTHS from the mailing date of this communication.
- If NO period for reply is specified above, the maximum statutory period will apply and will expire SIX (6) MONTHS from the mailing date of this communication.
- Failure to reply within the set or extended period for reply will, by statute, cause the application to become ABANDONED (35 U.S.C. § 133). Any reply received by the Office later than three months after the mailing date of this communication, even if timely filed, may reduce any earned patent term adjustment. See 37 CFR 1.704(b).

Status

- 1) ☒ Responsive to communication(s) filed on 21 November 2008.
- 2a) ☒ This action is **FINAL**. 2b) ☐ This action is non-final.
- 3) ☐ Since this application is in condition for allowance except for formal matters, prosecution as to the merits is closed in accordance with the practice under *Ex parte Quayle*, 1935 C.D. 11, 453 O.G. 213.

Disposition of Claims

- 4) ☒ Claim(s) 1-15 and 19-24 is/are pending in the application.
- 4a) Of the above claim(s) 6, 11, 12, 14, 15 and 19-22 is/are withdrawn from consideration.
- 5) ☐ Claim(s) _____ is/are allowed.
- 6) ☒ Claim(s) 1-5, 7-10 and 13-24 is/are rejected.
- 7) ☒ Claim(s) 5 is/are objected to.
- 8) ☐ Claim(s) _____ are subject to restriction and/or election requirement.

Application Papers

- 9) ☐ The specification is objected to by the Examiner.
- 10) ☒ The drawing(s) filed on 19 May 2005 is/are: a) ☒ accepted or b) ☐ objected to by the Examiner.
Applicant may not request that any objection to the drawing(s) be held in abeyance. See 37 CFR 1.85(a).
Replacement drawing sheet(s) including the correction is required if the drawing(s) is objected to. See 37 CFR 1.121(d).
- 11) ☐ The oath or declaration is objected to by the Examiner. Note the attached Office Action or form PTO-152.

Priority under 35 U.S.C. § 119

- 12) ☒ Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f).
- a) ☒ All b) ☐ Some * c) ☐ None of:
1. ☒ Certified copies of the priority documents have been received.
 2. ☐ Certified copies of the priority documents have been received in Application No. _____.
 3. ☐ Copies of the certified copies of the priority documents have been received in this National Stage application from the International Bureau (PCT Rule 17.2(a)).

* See the attached detailed Office action for a list of the certified copies not received.

Attachment(s)

- | | |
|--|---|
| 1) <input type="checkbox"/> Notice of References Cited (PTO-892) | 4) <input type="checkbox"/> Interview Summary (PTO-413) |
| 2) <input type="checkbox"/> Notice of Draftsperson's Patent Drawing Review (PTO-948) | Paper No(s)/Mail Date. _____ |
| 3) <input type="checkbox"/> Information Disclosure Statement(s) (PTO/SB/08) | 5) <input type="checkbox"/> Notice of Informal Patent Application |
| Paper No(s)/Mail Date _____ | 6) <input type="checkbox"/> Other: _____ |

DETAILED ACTION

Claim status

This action is in response to papers filed 11/21/2008.

Claims 1-15, 19-24 are pending.

Claims 6, 11-12, 15, 19-22 are withdrawn.

Claims 1-5, 7-10, 13, 23-24 are being examined.

The 102 based on Huang has been withdrawn in view of the amendment to delete, "complementary sequences thereof." The primers of Huang do not comprise primers of SEQ ID NO 76 and 77 and thus does not anticipate the claims.

The 103 based on Huang in view of Voelker, Huang in view of Hogan and Hopewell, Huang in view of Voelker and Southern have been withdrawn in view of the amendment to delete, "complementary sequences thereof." The primers of Huang do not comprise primers of SEQ ID NO 76 and 77 and thus does not render the claims obvious.

Claim Objections-Maintained

This objection has been maintained.

Claim 5 is objected to because it specifically recites nonelected subject matter. The claims require "a combination of oligonucleotide probes comprises all or a portion of the sequences identified with SEQ ID No 1 to 69". As stated in the response to the restriction filed 11/17/2006, applicant has elected a specific combination of SEQ ID NO 24. Applicant should amend the claims so that the claims are directed to the elected invention of the specific combination of genes.

Prior to allowance of these claims, the non-elected subject matter will be required to be deleted from the claims.

Response to Arguments

The response has requested the objection to claim 5 be held in abeyance until allowable matter be determined. Thus as no arguments have been presented this objection is maintained.

Claim Rejections - 35 USC § 112-Maintained

1. The following is a quotation of the second paragraph of 35 U.S.C. 112:

The specification shall conclude with one or more claims particularly pointing out and distinctly claiming the subject matter which the applicant regards as his invention.

2. Claims 1-5, 7-10, 13-14, 23-24 rejected under 35 U.S.C. 112, second paragraph, as being indefinite for failing to particularly point out and distinctly claim the subject matter which applicant regards as the invention.

Claim 1 recites the limitation " said detection " in step c. There is insufficient antecedent basis "said detection" for this limitation in the claim. This rejection can easily be overcome by amending the claim to recite, "detection of said hybridization complex."

Response to Arguments

This rejection is maintained as the claim 1 still recites "said detection." The response asserts the claim has been amended as suggested by the examiner, however the examiner suggested amending the claim to recite "detection of said hybridization complex" not "said detection of said hybridization complex." The examiner apologizes for any confusion.

Claim Rejections - 35 USC § 103-maintained

3. The following is a quotation of 35 U.S.C. 103(a) which forms the basis for all obviousness rejections set forth in this Office action:

(a) A patent may not be obtained though the invention is not identically disclosed or described as set forth in section 102 of this title, if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a person having ordinary skill in the art to which said subject matter pertains. Patentability shall not be negated by the manner in which the invention was made.

4. Claims 1-5, 7-10, 13, 23-24 are rejected under 35 U.S.C. 103(a) as being unpatentable over Haung (US Patent 5,645,994, Issued 1997) in view of Haselbeck et al (WO01/70955, published September 27, 2001).

It is noted that claim 5 recites, "oligonucleotide probes comprise all or a portion of sequences identified with SEQ ID NO 1 to 69." This recitation is being given the broadest reasonable interpretation that the claim requires a portion of the elected SEQ ID NO 24, which includes a single nucleic acid.

With regards to claim 1, Huang teaches a method of identifying species of bacteria in a sample by amplification with universal primers based on consensus amino acid sequences which flank variable amino acid sequences (see abstract). Haung further teaches a method of designing universal primer that amplify parE and gyrB (see column 6 lines 28-65). Haung further teaches the use of universal primer compositions to amplify gyrB and parE sequences (see column 14, lines 16-19). Haung further teaches the use of nested primers to specifically distinguish between closely related species (see column 15 lines 27-35). The nested probes are thus the equivalents of the probes claimed. Haung further teaches, "In theory, a single pair of primers, one from each flanking consensus sequence, can be used to amplify the signature sequence.

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However, a highly preferred embodiment includes a multiplicity of primers having sequences corresponding to potential alternate DNA sequences. As is well-known, the genetic code is degenerate, meaning that an individual amino acid may be coded for by as many as 6 different DNA codons (each codon consisting of three adjacent nucleotides). Thus, even though the amino acid sequence of a region of type II topoisomerase from different organisms may be identical, the DNA in those organisms which codes for the region may differ. The PCR technique requires a good match between the DNA primer sequences especially at the 3' end and the DNA to which it binds (Saiki et al.). Thus, to avoid failing to amplify species having such alternate DNA sequences, the set of primers should include variant primers having at least some of the alternate sequences. Moreover, it is desirable that the amino acids in the consensus sequence be coded for by 3 or fewer different codons, especially in the portion immediately adjacent to the signature segment. Obviously, the presence of one or more amino acids having six possible codons drastically increases the number of possible DNA sequences. By choosing the consensus sequences to have amino acids with at most three possible codons (or in an even more preferred embodiment, two possible codons), the number of different oligonucleotide sequences required in the universal primers is kept manageable" (see column 7, line 60 to column 8, line 9). Huang teaches, "It will be recognized that universal primers such as the compositions described herein can be constructed for any ubiquitous protein having substantially conserved segments adjacent to variable segments. Depending upon the desired application, gene products other than type II topoisomerase might be preferable.

Examples of proteins of potential use according to this invention include RNA polymerase and other DNA binding proteins. Where it is desired only to distinguish among very closely related species, a protein common only to such species may be used" (see column 14, lines 46-50). Huang concludes, "it will be apparent how a specific primer pair for any species can be designed by the methods disclosed herein and using a database" (see column 16, lines 50-55).

With regards to claim 2, Haung further teaches the use of universal primer compositions to amplify *gyrB* and *parE* sequences due to their sequence similarities (see column 14, lines 16-19). Haung further teaches identification of *legionella pneumophila* (SEQ ID NO 70), which is a bacteria that infects the respiratory tract.

With regards to claim 3, Huang teaches detection of *pseudomonase aeruginosa* (see figure 9 a).

With regards to claim 4, Haung et al teaches the use of primers of 15 to 36 nucleotides in length (see column 7, lines 22-25).

With regards to claims 23 and 24 Huang et al teaches primers of 24 bases (see SEQ ID NO 206 and SEQ ID NO 207).

Huang does not teach primers comprising SEQ ID NO 76 and 77.

However, Haselbeck et al teaches SEQ ID NO 9244, which contains nucleotides 82 to 101 that comprise SEQ ID NO 76 of instant claims. Haselbeck further teaches SEQ ID NO 1844, which contains nucleotides 14 to 33 which comprise SEQ ID NO 77. Haselbeck teaches, "The identified or isolated nucleic acids obtained using the PCR primers may contain part or all of the homologous nucleic acids. Because homologous

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nucleic acids are related but not identical in 25 sequence, those skilled in the art will often employ degenerate sequence PCR primers. Such degenerate sequence primers are designed based on sequence regions that are either known to be conserved or suspected to be conserved such as conserved coding regions. The successful production of a PCR product using degenerate probes generated from the sequences identified herein would indicate the presence of a homologous gene sequence in the species being screened. The PCR primers 30 are at least 10 nucleotides, and preferably at least 20 nucleotides in length. More preferably, the PCR primers are at least 20-30 nucleotides in length. In some embodiments, the PCR primers can be more than 30 nucleotides in length. It is preferred that the primer pairs have approximately the same G/C ratio, so that melting temperatures are approximately the same.” (see page 232, lines 23-33). Haselbeck teaches multiple nucleic acid sequences that comprise a multiple nucleotides that are broadly interpreted as a portion of SEQ ID No 24.

With regards to claims 7-9, Haselbeck teaches, “In another embodiment, gene expression arrays and microarrays can be employed. Gene expression arrays are high density arrays of DNA samples deposited at specific locations on a glass chip, nylon membrane, or the like. Such arrays can be used by researchers to quantify relative gene expression under different conditions. Gene expression arrays are used by researchers to help identify optimal drug targets, profile new compounds, and determine disease pathway” (see page 78, lines 1-5).

With regards to claim 10, Haselbeck teaches use of a single stranded labeled probe (see page 231, lines 5-20).

With regards to claim 10, Haselbeck teaches the use of nucleic acid probes to identify microorganism species from clinical specimens (see page 195, line 20). Haselbeck further teaches, "Single stranded labeled cDNAs are prepared for hybridization to the array" (see page 149, line 17)

Therefore it would have been prima facie obvious to one of ordinary skill in the art at the time the invention was made to combine the method of Huang with the nucleic acids taught by Haselbeck which comprise SEQ ID NO 76 and 77. The skilled artisan would be motivated to combine the teachings of Huang and Haselbeck because Huang suggests the use of his method with any protein and any sequence in a database for identification of bacteria. The skilled artisan would also be motivated to combine the teachings of Huang and Haselbeck because Haselbeck suggests the use of his method to identify microorganism species.

It would further have been prima facie obvious to one of ordinary skill in the art at the time the invention was made to use arrays or microarrays in the combined method of Huang and Haselbeck because Haselbeck teaches it allows the detection of multiple gene expression products and thus markers.

The artisan would have a reasonable expectation of success in combining the teachings of Huang and Haselbeck as both teach methods of identifying microorganism by hybridization using methods and techniques known in the art.

Response to arguments

The response correctly notes on page 8 that Huang does not teach primers comprising SEQ ID NO 76 and SEQ ID NO 77, as noted in the instant rejection. Haselbeck is being used for those teachings.

The response further asserts that the instant method is faster and more reliable than those methods in the prior art and directs the examiner to example 6. Example 6 of instant specification teaches primers directed to 16s rRNA which are different than the instant invention or the prior art of Huang and Haselbeck. Thus example 6 does not suggest that the instantly claimed method is more sensitive or specific than the combination of Haselbeck and Huang. It is noted that example 6 refers to prior art references by Nikkari and Kotilainen, which appear to be the 16s rRNA studies of comparison, however these references have not been provided or cited on an IDS and thus have not been considered other than their description in example 6.

The response further asserts that the instant invention allows for multiplex analysis to distinguish between desired bacteria and normal flora as exemplified in example 7. These arguments have been thoroughly reviewed but are not considered persuasive as the example 7 of the specification comparison again appears to a method using 16s rRNA as described by the prior art of Hendolin, which has not been provided or presented on an IDS. Thus the example 7 of the specification does not teach or suggest the claimed invention has unexpected results differentiating it from Huang and Hselbeck.

First, MPEP 716.01(c) makes clear that "The arguments of counsel cannot take the place of evidence in the record. In re Schulze , 346 F.2d 600, 602, 145 USPQ 716,

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718 (CCPA 1965). Examples of attorney statements which are not evidence and which must be supported by an appropriate affidavit or declaration include statements regarding unexpected results, commercial success, solution of a long - felt need, inoperability of the prior art, invention before the date of the reference, and allegations that the author(s) of the prior art derived the disclosed subject matter from the applicant." Here, the statements regarding the improved sensitivity and specificity must be supported by evidence, not argument as described above examples 6 and 7 of the instant specification do not provide evidence as they are comparison to degenerate primers to the 16s rRNA genes, not the topoisomerase genes of the instant claims and thus do not provide evidence.

This should not be construed as an invitation for providing evidence. As further stated in the MPEP 716.01 regarding the timely submission of evidence:

A) Timeliness.

Evidence traversing rejections must be timely or seasonably filed to be entered and entitled to consideration. In re Rothermel, 276 F.2d 393, 125 USPQ 328 (CCPA 1960). Affidavits and declarations submitted under 37 CFR 1.132 and other evidence traversing rejections are considered timely if submitted:

- (1) prior to a final rejection,
- (2) before appeal in an application not having a final rejection, or
- (3) after final rejection and submitted
 - (i) with a first reply after final rejection for the purpose of overcoming a new ground of rejection or requirement made in the final rejection, or

- (ii) with a satisfactory showing under 37 CFR 1.116(b) or 37 CFR 1.195, or
- (iii) under 37 CFR 1.129(a).

The response continues by acknowledging that methods of designing universal primers and probes as well as microarray technology have been known for years. The response continues by asserting that in silico design methods of primers and probes does not necessarily correspond to wet-lab behavior of primers and probes. The response further asserts that multiple experiments would be required to find probes with superior behavior. The response presents Wiesinger-Mayr as evidence of this and quotes page 2 probe and array design. These arguments have been thoroughly reviewed but are not considered persuasive first as again the teachings of Wiesinger-Mayr are drawn to the 16s rRNA, not the claimed topoisomerase genes. Wiesinger-Mayr teaches, "An in silico hybridisation matrix was generated with the Probe Match function in the ARB software package and the CalcOligo software. The modeled hybridization behavior of each probe was in good agreement with real experimental data" (page 4, bottom 1st column, top 2nd column). Further, Wiesinger-Mayr teaches, "Due to the application of short oligonucleotide probes it was possible to discriminate sequences differing from each other by a single nucleotide" (page 7, 1st column, top). Wiesinger-Mayr in the discussion teaches, "Furthermore the use of the most characterized marker gene with the highest number of available sequences allows reliable in silico predictions based on experimentally obtained data" (page 10, 2nd column, 1st full paragraph). Thus when taken as a whole Wiesinger-Mayr does not teach away from in silico design, but

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suggest it is predictable. Further any experimentation required for testing of the primers or probes would be routine, absent secondary considerations in view of the teachings of Wiesinger-Mayr.

The response again asserts that one of skill in the art would not be able to predict the superior results of the particular primers recited in the claim. This argument has been thoroughly reviewed but is not considered persuasive as the claims are drawn to primers comprising the sequences of SEQ ID NO 76 and 77 and Hasebelbeck teaches sequences comprising SEQ ID NO 76 and 77. The comprising language of the claim allows for the inclusion of additional nucleotides, while the specification does not provide a definition limiting the length of the primers. Further the response has asserted the instant primers have superior results, but has not provided evidence to support such an assertion. First, MPEP 716.01(c) makes clear that "The arguments of counsel cannot take the place of evidence in the record. In re Schulze , 346 F.2d 600, 602, 145 USPQ 716, 718 (CCPA 1965). Examples of attorney statements which are not evidence and which must be supported by an appropriate affidavit or declaration include statements regarding unexpected results, commercial success, solution of a long - felt need, inoperability of the prior art, invention before the date of the reference, and allegations that the author(s) of the prior art derived the disclosed subject matter from the applicant." Here, the statements regarding superior results must be supported by evidence, not argument.

This should not be construed as an invitation for providing evidence. As further stated in the MPEP 716.01 regarding the timely submission of evidence:

A) Timeliness.

Evidence traversing rejections must be timely or seasonably filed to be entered and entitled to consideration. In re Rothermel, 276 F.2d 393, 125 USPQ 328 (CCPA 1960). Affidavits and declarations submitted under 37 CFR 1.132 and other evidence traversing rejections are considered timely if submitted:

- (1) prior to a final rejection,
- (2) before appeal in an application not having a final rejection, or
- (3) after final rejection and submitted
 - (i) with a first reply after final rejection for the purpose of overcoming a new ground of rejection or requirement made in the final rejection, or
 - (ii) with a satisfactory showing under 37 CFR 1.116(b) or 37 CFR 1.195, or
 - (iii) under 37 CFR 1.129(a).

The response continues by noting that applicant's have aligned multiple bacterial sequences to the primers of SEQ ID NO 76 and 77 of the instant claims as well as those of Huang. The arguments to the primers of Huang are moot as Haselbeck is being relied upon for teaching sequences comprising SEQ ID NO 76 and 77.

The response then asserts that the inventors had to take into account cross reaction with all probes in the panel as they are all present on the microarray. First examiner notes that applicant elected the combination of one probe with respect to claim 5 and claim 1 is generic to the combination. Thus the claims in view of the election encompass a single probe and cross reactivity is not an issue when only one

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probe is required. Further Haselbeck is being used for the teachings of a probe comprising all or a portion of SEQ ID NO 24 and/or complementary sequences thereof and thus the combination of Huang and Haselbeck render the instant claims obvious.

The arguments to Huang not leading to high throughput array application have been thoroughly reviewed but are not considered persuasive as the applicant elected the combination of SEQ ID NO 24, thus the claims require a combination of 1 probe and not the multiple probes asserted. Further Huang is being used in combination with Haselbeck which does suggest the use of microarrays, thus making a broader interpretation of the claims obvious as well.

The response conclude by asserting that while the longer sequences were known the instant claimed method uses shorter probes and primers that are a significant improvement over the art. These arguments have been thoroughly reviewed but are not considered persuasive as addressed previously the response has provided no evidence of the asserted "significant improvement" by comparison to methods in which topoisomerase genes were examined. Further the claims are drawn to comprising language and thus encompass not the short primers consisting of SEQ ID NO 76 and SEQ ID NO 77 and a short probe consisting of SEQ ID NO 24, but also longer primers and probes. Further claim 5 also encompasses, "all or a portion of the sequences of SEQ ID NO 24, and/or complementary sequences thereof." Thus claim 5 broadly encompasses any fragment of any length of SEQ ID NO 5.

Thus the rejection is maintained in view of the arguments above.

5. Claim 5 is rejected under 35 U.S.C. 103(a) as being unpatentable over Haung (US Patent 5,645,994, Issued 1997) in view of Haselbeck et al (WO01/70955, published September 27, 2001) as applied to claims 1-5, 7-10, 13, 23-24 above, and further in view of Hogan et al (US Patent 5541308) and Hopewell et al (Journal of Bacteriology (1990), volume 172, pages 3481-3484).

This rejection is drawn to the interpretation claim 5 requires a probe comprising the entire elected sequence of SEQ ID NO 24 or a sequence that is fully complementary to SEQ ID NO 24.

The teachings of Huang and Haselbeck are set forth in paragraph 4 above.

Haung and Haselbeck do not teach the probe of the comprising all or a portion of SEQ ID NO 24.

However, Hogan et al teaches probe design for detection of specific sequences (see abstract). Hogan teaches identification of variable regions (see column 6, lines 3-55). Hogan teaches alignment of these variable regions (see column 6 line 67—column 7, line 8). Hogan further teaches probes should be positioned to minimize stability of probe:nontarget hybrids, by avoiding GC rich regions and areas of frequent mutation (see column 7 lines 10-15). Hogan teaches the use of synthetic oligonucleotide probes of 15-50 base pairs (see column 10, lines 40-45). Hogan further teaches maximizing stability of probe target hybrid, by avoiding long AT sequences and terminating hybrids with G:C base pairing and the appropriate T_m (see column 7 lines 16-19). Hogan further teaches targeting sequences known to have secondary structure issues and probes that are self-complementary should be avoided (see column 7, lines 20-29).

Hopewell teaches sequence of *Staphylococcus aureus* gyrB, which comprises SEQ ID NO 24, (see figure 3B). Hopewell teaches that quinolone resistant *Staphylococcus aureus* are a major medical problem and this resistance is due to mutations in the DNA gyrase enzyme (see page 3481, 1st column, 1st paragraph).

Designing probes, which are equivalents to those taught in the art is routine experimentation. The prior art teaches the parameters and objectives involved in the selection of oligonucleotides that function as probes, see Hogan. Moreover there are many internet web sites that provide free downloadable software to aid in the selection of probes drawn from genetic data recorded in a spreadsheet. The prior art is replete with guidance and information necessary to permit the ordinary artisan in the field of nucleic acid detection to design probes. As discussed above, the ordinary artisan would be motivated to have designed and tested new probes to obtain additional oligonucleotides that function to detect specific hypervariable regions of bacteria and identify oligonucleotides with improved properties. The ordinary artisan would have a reasonable expectation of success of obtaining additional probes from within the sequences provided by Haselbeck using the method described by Hogan and Huang. Thus, for the reasons provided above, the ordinary artisan would have designed additional probes using the teachings in the art at the time the invention was made. The claimed SEQ ID NOs are obvious over the cited prior art, absent secondary considerations.

Therefore it would have been prima facie obvious to one of ordinary skill in the art at the time the invention was made to use the sequence taught by Hopewell and the

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probe design method of Hogan to make probes to detect bacterial species based on the gyrB. The ordinary artisan would thus design a probe comprising SEQ ID NO 24 or complementary to SEQ ID NO 24. The ordinary artisan would be motivated to use the sequence taught by Hopewell to design probes by Hogan's method of probe design to identify mutations that result in quinolone resistance because Hopewell teaches this is a serious medical problem and proper identification would allow efficient treatment. The artisan would have a reasonable expectation of success as the methods are drawn to well known methods of making and analyzing nucleic acids known in the art.

Response to arguments

The response has provided no specific arguments to the combination of Huang, Haselbeck, Hogan and Hopewell. The arguments directed to independent claim 1 has been addressed above. Thus as there is no arguments to the obviousness of claim 5 in view of Huang, Haselbeck, Hogan and Hopewell the rejection is maintained.

Summary

No claims are allowed over prior art cited.

Conclusions

6. Applicant's amendment necessitated the new ground(s) of rejection presented in this Office action. Accordingly, **THIS ACTION IS MADE FINAL**. See MPEP § 706.07(a). Applicant is reminded of the extension of time policy as set forth in 37 CFR 1.136(a).

A shortened statutory period for reply to this final action is set to expire THREE MONTHS from the mailing date of this action. In the event a first reply is filed within TWO MONTHS of the mailing date of this final action and the advisory action is not mailed until after the end of the THREE-MONTH shortened statutory period, then the shortened statutory period will expire on the date the advisory action is mailed, and any extension fee pursuant to 37 CFR 1.136(a) will be calculated from the mailing date of the advisory action. In no event, however, will the statutory period for reply expire later than SIX MONTHS from the date of this final action.

Any inquiry concerning this communication or earlier communications from the examiner should be directed to Steven C. Pohnert whose telephone number is (571)272-3803. The examiner can normally be reached on Monday-Friday 6:30-4:00, every second Friday off.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Ram Shukla can be reached on 571-272-0735. The fax phone number for the organization where this application or proceeding is assigned is 571-273-8300.

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Information regarding the status of an application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or Public PAIR. Status information for unpublished applications is available through Private PAIR only. For more information about the PAIR system, see <http://pair-direct.uspto.gov>. Should you have questions on access to the Private PAIR system, contact the Electronic Business Center (EBC) at 866-217-9197 (toll-free). If you would like assistance from a USPTO Customer Service Representative or access to the automated information system, call 800-786-9199 (IN USA OR CANADA) or 571-272-1000.

Steven Pohnert

**/Jeanine A Goldberg/
Primary Examiner, Art Unit 1634**